Gene Flow Between Chromosomal Forms of the Malaria Vector *Anopheles funestus* in Cameroon, Central Africa, and Its Relevance in Malaria Fighting

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ABSTRACT

Knowledge of population structure in a major vector species is fundamental to an understanding of malaria epidemiology and becomes crucial in the context of genetic control strategies that are being developed. Despite its epidemiological importance, the major African malaria vector *Anopheles funestus* has received far less attention than members of the *Anopheles gambiae* complex. Previous chromosomal data have shown a high degree of structuring within populations from West Africa and have led to the characterization of two chromosomal forms, “Kiribina” and “Folonzo.” In Central Africa, few data were available. We thus undertook assessment of genetic structure of *An. funestus* populations from Cameroon using chromosomal inversions and microsatellite markers. Microsatellite markers revealed no particular departure from panmixia within each local population and a genetic structure consistent with isolation by distance. However, cytogenetic studies demonstrated high levels of chromosomal heterogeneity, both within and between populations. Distribution of chromosomal inversions was not random and a cline of frequency was observed, according to ecotypic conditions. Strong deficiency of heterokaryotypes was found in certain localities in the transition area, indicating a subdivision of *An. funestus* in chromosomal forms. *An. funestus* microsatellite genetic markers located within the breakpoints of inversions are not differentiated in populations, whereas in *An. gambiae* inversions can affect gene flow at marker loci. These results are relevant to strategies for control of malaria by introduction of transgenes into populations of vectors.

MALARIA is a devastating disease killing >1.5 million people every year, mostly among African children under the age of 5 (World Health Organization 1993). Although efficient means for control do exist, their implementation in African settings has so far remained ineffective and the burden of the disease is still increasing (Greenwood and Mutabingwa 2002). Innovative strategies to combat malaria are thus needed. Plasmodium parasites, the causative agents of malaria, are transmitted from human to human through the bite of a mosquito vector that belongs to the genus *Anopheles*. Genetic control of wild vector populations to make them unable to transmit the parasite has therefore been proposed as a way to break down malaria transmission. The idea has gathered considerable research momentum during the past decades. Owing to recent development in molecular entomology, bioinformatics, and genomics, this view is currently considered one of the most promising approaches to bring malaria transmission under control (James et al. 1999; Collins et al. 2000; Alphey et al. 2002). Several important goals for the development of transgenic mosquitoes have indeed already been achieved, including germ-line transformation of *Anopheles* (Catteruccia et al. 2000; Grossman et al. 2001) and identification of several “candidate” genes involved in refractoriness of *Anopheles* to Plasmodium (Shahabuddin et al. 1998; Ito et al. 2002; Blandin et al. 2004; Osta et al. 2004). A number of genetic constructs are being considered as potential genetic drive systems that would facilitate the spread of effector genes into target vector populations and eventually lead to complete replacement of wild populations (Kidwell and Ribeiro 1992; Ribeiro and Kidwell 1994; Turelli and Hoffmann 1999; Sinkins and O’Neill 2000). However, availability of these tools does not imply success of such a campaign. A comprehensive knowledge of the population genetic structure of the target species and of the forces that generate and maintain this structure...
is required to assess feasibility of this strategy, because
the rate of gene flow in a subdivided population can be
prohibitively slow (Lanzaro and Tripet 2003). Current
vector control efforts involving insecticides will also ben-
efit from knowledge of gene flow, which allows predic-
tion of the spread of genes conferring insecticide resis-
tance within and between natural vector populations.

To date, most population genetics studies targeting
malaria vector species have focused on Anopheles gam-
biae, the major human malaria vector in sub-Saharan
Africa, and demonstrated highly complex population
 genetic structure (Donnelly et al. 2002; see Tripet et
al. 2004, accompanying article in this issue). However,
a number of highly efficient malaria vectors occur in
the field, but these have received far less attention than
members of the An. gambiae complex, despite their epi-
demiological importance (Fontenille and Simard
2004). This is the case of the widespread Anopheles fu-
nestus, which has a high vectorial ability, sometimes
higher than An. gambiae (Fontenille et al. 1997; Manga
et al. 1997). Vector control strategies must therefore
also consider An. funestus as a main target species. In
West Africa, An. funestus populations showed a high level
of genetic and behavioral heterogeneity. Cyto genetic
studies conducted in Burkina Faso and Senegal revealed
highly significant departures from Hardy-Weinberg
equilibrium for most chromosomal paracentric inver-
sion systems and linkage disequilibrium between inver-
sions located on different chromosomes, suggesting that
An. funestus populations could be divided in chromo-
somal forms with low or no gene flow between them
(Lochouarn et al. 1998; Costantini et al. 1999; Dia
et al. 2000a). Paracentric chromosomal inversions are
known to reduce recombination between alternative ar-
rangements (Sturtevant and Beadle 1936), especially
at and close to the breakpoints (Navarro et al. 1997;
Andolfatto et al. 2001). As such, they are commonly
viewed as adaptive mechanisms that can capture and
stabilize blocks of coadapted genes with strong pheno-
typic effect, hence allowing ecological expansion in mar-
ginal habitats (Mayr 1963; Dobzhansky 1970). Ecologi-
cal segregation of carriers of different inversion, coupled
with strong genetic drift in isolated populations, could
then lead to parallel segregation of mating recognition
variants and speciation. A number of inversion-based
speciation models have indeed been suggested (Coluzzi
1982; King 1993; Noor et al. 2001; Reeserberg 2001). Two
chromosomal forms were defined for An. funestus in
Burkina Faso: the first one, “Kiribina,” was characterized
by standard chromosomes or presence of inversion 2Rs;
the second one, “Folonzo,” was mainly polymorphic on
chromosomal arm 2 and presented high frequencies of
inversions 3Ra, 3Rb, and/or 3La (Costantini et al.
1999; Gueulebeogo et al. 2002). In South and East Africa,
however (namely, Angola and Kenya), chromosomal
inversions were found polymorphic but, contrary to
West Africa, heterokaryotypes were found at the ex-
pected Hardy-Weinberg frequencies in each location
and no evidence for population subdivision was found
(Sharakhov et al. 2001; Boccolini et al. 2002; Kamau
et al. 2003). Thus, chromosomal structuring of wild An.
funestus populations appears highly variable throughout
Africa.

Very few data from Central Africa are available to date.
A handful of populations from South Cameroon were
analyzed for chromosomal inversion polymorphisms and
fall within the definition of the Folonzo form, with addi-
tional inversions 2Rd and 2Rh that are commonly found
in East Africa but absent from West African populations
(Dia et al. 2000b). Taking advantage of the central geo-
ographic position of Cameroon in Africa, of the extreme
diversity of ecological settings and widespread distribu-
tion of An. funestus throughout the country, we under-
took a study of the genetic structure of the species by
sampling An. funestus populations along a north-south
transect in Cameroon representing a clear cline in arid-
ity (Figure 1). Genetic diversity and population differen-
tiation was assessed using cyto genetics (i.e., chromo-
somal inversions distribution) and genotyping at 10
microsatellite markers that map both within and outside
inversions (Sharakhov et al. 2004). This study design
allowed us to explore the extent of genetic structuring
within and between natural populations of this highly
efficient malaria vector.

MATERIALS AND METHODS

Sampling sites: Mosquitoes were collected across Cameroon
along a transect from south to north: in Mfou (3°41′ N, 11°32′
E), Mbebe (4°19′ N, 11°32′ E), Ntui (4°26′ N, 11°37′ E),
Nkoteng (4°30′ N, 12°03′ E), Foumban (5°47′ N, 10°43′ E),
Bankim (6°20′ N, 11°22′ E), Tibati (6°28′ N, 12°37′ E),
Ngaoundere (7°19′ N, 13°35′ E), Lagdo (9°30′ N, 13°44′ E),
and Maga (10°51′ N, 14°57′ E) (Figure 1).

Humid forest in the south of the country gradually turns
into arid savannas in the north. Cameroon can roughly be
divided into four climatic zones, based on amount and distri-
bution of annual rainfall (Olivry 1986). According to this
classification, Mfou, Mbebe, Ntui, Foumban, and Bankim lie
within the equatorial climate zone, with mean annual rainfalls
ranging from 1600 to 1800 mm. In this area two rainy seasons
occur: from March to June and from September to November.
Tibati, Ngaoundere, and Maroua are located in the Sudanian
climatic domain. Mean annual rainfall in this area decreases
from south to north, with 1700 mm in Tibati and 900 mm in
Lagdo. Only one rainy season occurs during the year, the
number of rainy months decreasing from south to north. In
Tibati the rainy season extends from March to November and
in Lagdo from May to October. Maga is in the Sudano-Sahelian
climatic domain, with mean annual rainfalls ~750 mm and
one rainy season from May to September.

Mosquito collection and field processing of specimens:
Specimens were collected between October 2000 and January
2003 either after landing on human volunteers or by indoor
pyrethrum spraying. Specimens from Mfou, Mbebe, and
Nkoteng were collected during the rainy season in May 2001;
specimens from Ngaoundere, Lagdo, and Maga were collected
during the dry season in December 2001 and specimens from
Ntui, Foumban, Bankim during the dry season in January
Gene Flow in *A. funestus*

2003. Specimens from Tibati were collected during the rainy seasons of 2000 and 2002. Females from the *An. funestus* group were visually sorted from other anophelines according to morphological identification keys (Gillies and de Meillon 1968; Gillies and Coetzee 1987). All specimens were stored individually in tubes containing desiccant. Half-gravid females of the *An. funestus* group were dissected and ovaries, which contain polytene chromosomes on which chromosomal inversions can be scored, were immediately fixed in ice-cold Carnoy’s fixative (one part of glacial acetic acid and three parts of pure ethanol). The corresponding carcasses were stored in tubes with desiccant. Back in the laboratory, all tubes were stored at −20°C until processed.

**DNA extraction and species identification:** Genomic DNA was extracted from wings and legs of each individual mosquito following a slightly modified version of the protocol of Cornel and Collins (1996) and resuspended in sterile water. *An. funestus* s.s. females were included in the analysis after species identification was carried out by the recently described diagnostic PCR assay (Koekemoer et al. 2002; Cohuet et al. 2003). No other member of the *An. funestus* group was observed in our samples.

**Microsatellite amplification:** Ten microsatellites loci were selected from published *An. funestus* sequence data (Sinkins et al. 2000; Cohuet et al. 2002) on the basis of high polymorphism, apparent absence of null alleles, mean allele sizes ranging from 110 to 220 bp, and cytological location allowing whole-genome scan (Figure 2). We used AFND5 and FunO located on chromosomal arm 2R; FunL on chromosomal arm 3L; FunG, FunD, AFND19, and AFND20 on chromosomal arm 3R; FunF on chromosomal arm 3L; and AFND2 and AFND3, whose precise cytological location is still undefined (Sharakhov et al. 2004). PCR amplification was carried out in 25 μl reaction volume from 5 to 10 ng of template DNA. Reaction mixture contained 1× PCR buffer containing 1.5 mM MgCl₂ (QIAGEN, Courtaboeuf, France), 200 μM each dNTP, 10 pmol of each primer, and 0.5–1 unit Taq Polymerase (QIAGEN). The forward primer was labeled in 5’ with TET, HEX, or FAM fluorescent markers (Eurogentec, Seraing, Belgium) to allow multiplex electrophoresis. Amplification was performed under the following conditions: an initial denaturation step at 94°C for 2 min followed by 36 cycles of 30 sec at 94°C, 50 sec at 54°C, 30 sec at 72°C, and a final elongation step of 10 min at 72°C. Fragment analyses were conducted with an ABI PRISM 377. Alleles were sized relatively to an internal size standard using GENESCAN version 3.1 (Applied Biosystems, Foster City, CA).

**Polytene chromosome preparation:** Ovaries of half-gravid females were used to obtain squash preparations of the polytene chromosomes according to Hunt (1973). The preparations were examined under phase-contrast microscopy, and paracentric inversion karyotypes were scored according to the chromosomal map and nomenclature of Sharakhov et al. (2004).

**Data analysis:** For each microsatellite locus, deviation from Hardy-Weinberg expectations was tested in each location and overall. Cytogenetic data (i.e., karyotypes) were analyzed in the same way, considering alternative chromosomal arrangements as different alleles at one locus as previously defined (Lochouarn et al. 1998; Costantini et al. 1999; Dia et al. 2000a,b). Statistical significance was assessed by the exact probability tests available in GENEPOP 3.2 (Raymond and Rousset 1995). Statistical significance for linkage disequilibrium between pairs of microsatellite loci in the pooled population and within each population were computed by exact tests using GENEPOP 3.2. For specimens analyzed with both chromosomal and microsatellite markers, cytogenetic data were included in the analysis.

Genetic differentiation between geographical populations was examined by F statistic (Wright 1951), calculated according to Weir and Cockerham (1984). Significance of Fst was assessed using the G-based exact test of genotypic differentiation (Goudet et al. 1996). The correlation between genetic and geographical distances was assessed by the regression of Fst/(1 – Fst) on the logarithm (ln) of geographical distance (Rousset 1997) and tested using the Mantel test available in GENEPOP.

Multilocus and/or multisample tests of deviations from Hardy-Weinberg equilibrium were done using Fisher’s combination probabilities test using P values from each locus and/or population, as implemented in GENEPOP. Multilocus tests of population differentiation were conducted in the same way. The sequential Bonferroni procedure (Holm 1979) can detect a single test-specific departure when multiple tests are performed; it was applied to evaluate significance of locus-specific departures within populations.

**RESULTS**

**Chromosomal diversity and inversions distribution:** Cytological analysis was conducted in six villages: Ntui, Nkoteng, Bankim, Tibati, Ngaoundere, and Lagdo along the transect from south to north (Figure 1). All inversions found in Cameroon are schematized in Figure 2. No inversion was found on chromosomal arm 2L or on the X chromosome in accordence with former studies in this species. Polymorphism on chromosome arm 2R was low, with only three inversions observed (2Ra, 2Rd, and 2Rh). As a result of extensive overlapping and apparent sharing of breakpoints among inversions 2Ra, 2Rd, and 2Rh, these were never found to occur together on a single chromatid. We therefore treated them as different alleles of the same locus. Two independent inversions were observed on arm 3R (3Ra and 3Rb) and a single inversion was observed on arm 3L (3La).

All three specimens karyotyped in Nkoteng were homozygous for inversions 3Ra, 3Rb, and 3La. Inversions 2Rh and 2Rd were observed on arm 2R (Table 1). As 45 specimens previously analyzed in this village showed identical karyotypes, we could consider this population as fixed for inversions 3Ra, 3Rb, and 3La and that two inversions, 2Rh and 2Rd, exist on arm 2R (Dia et al. 2000b). Inversions 2Rh and 2Rd were found in Nkoteng, Ntui, and Tibati. Inversion 2Rd was always found at the heterozygote state with inversion 2Rh as previously recorded in this region (Dia et al. 2000b). Inversion 2Ra was observed in only two sampling sites (Ntui and Bankim). The frequency of the standard (2R+) arrangement increased in northern populations and was fixed beyond Ngaoundere. Inversions 3Ra and 3Rb were fixed in Nkoteng and absent in Lagdo where only the standard (3R+) arrangement was found. Clinal frequencies were found in localities in between. Inversion 3La was fixed in Ntui and Nkoteng and its frequency decreased in northern populations until becoming rare in Lagdo. Clinal distribution of chromosomal inversions was therefore observed on chromosomal arms 2R, 3R, and 3L, with inversions being more represented in villages from...
forested areas and standard arrangements more frequently found in drier savannas, with a gradient of frequencies along the latitudinal transect (Figure 1).

When considering all karyotyped specimens as belonging to a single population, Hardy-Weinberg equilibrium was significantly rejected ($P < 10^{-4}$) for all chromosomal inversions with a deficit of heterozygotes (Table 1, “Pooled”). All pairs of chromosomal inversions were in linkage disequilibrium (Table 2).

Following Costantini et al. (1999), all specimens from Nkoteng and Ntui belong to the Folonzo form while all those from Lagdo belong to Kiribina. Both chromosomal forms were found in Bankim, Tibati, and Ngaoundere, within the transitional zone between forest and savannas. In Tibati, where sample size was appropriate for subsequent analysis, significant departure from Hardy-Weinberg equilibrium was observed for all inversions due to a deficit of heterokaryotypes (Table 1). Hardy-Weinberg equilibrium was reestablished when individuals were assigned to chromosomal forms using the set of rules proposed by Costantini et al. (1999). Linkage disequilibrium between all pairs of inversions was highly significant ($P < 0.01$) in Tibati: $2Rd, 2Rh, 3Ra, 3Rb,$ and $3La$ were almost always associated while all those from Lagdo belong to Kiribina. Both together while arrangements $2Rd, 3Ra, 3Rb,$ and $3La$ occurred together in the same individuals, despite the fact that arms $2R$ and $3R$ are on two different chromosomes. This is consistent with a genetic subdivision of An. funestus populations in distinct chromosomal forms in this location. In Ngaoundere a similar pattern was observed, but $F_{IS}$ values were not statistically significant, probably because of low sample size in this locality. In contrast, in Bankim, Hardy-Weinberg expectations were not rejected, except for the inversion system on chromosomal arm $3L$, and no linkage disequilibrium was significant between inversions.

**Microsatellite diversity and neutral population structure**: Genotypes at 10 microsatellite loci were determined for all karyotyped and additional An. funestus specimens. In total, 583 adult females, collected in 10 villages spread along a south-north transect in Camer-
### TABLE 1

Inversion frequencies and significance level for goodness-of-fit tests to Hardy-Weinberg equilibrium for chromosomal inversions within *An. funestus* populations from Cameroon

<table>
<thead>
<tr>
<th>Inversion systems</th>
<th>Populations</th>
<th>Ntui</th>
<th>Nkoteng</th>
<th>Bankim</th>
<th>Tibati</th>
<th>Ngaoundere</th>
<th>Lagdo</th>
<th>Pooled*</th>
</tr>
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<tbody>
<tr>
<td>2Rdha</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
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<td></td>
</tr>
<tr>
<td>2n</td>
<td></td>
<td>12</td>
<td>6</td>
<td>14</td>
<td>46</td>
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<td>0</td>
<td>0</td>
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<td>F_\text{IS}</td>
<td></td>
<td>0.29*</td>
<td>-1</td>
<td>-0.09</td>
<td>0.66**</td>
<td>—</td>
<td>—</td>
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<td></td>
<td></td>
<td></td>
<td>0.74**</td>
</tr>
</tbody>
</table>

F_\text{IS} was calculated according to Weir and Cockerham (1984). 2n, number of chromatids scored. *P < 0.05; **P < 0.05 after Bonferroni correction (see text). —, sample size too low to warrant analyzes or only one arrangement found.

*All specimens were considered as belonging to one single panmictic population.

Gene Flow in *A. funestus*

Oon (Figure 1), were analyzed. Four microsatellites loci mapped within polymorphic chromosomal inversions: AF5 in 2Rd, 2Rh, and 2Ra, FunD and FunG in 3Rb, and FunF in 3La. Four other loci, FunO, FunL, AFND20, and AFND19, were located outside chromosomal inversions (Figure 2), while the precise cytological location of AF2 and AF3 is still unknown (Sharakhov et al. 2004). All loci were highly polymorphic, showing between 8 and 32 alleles per locus. Similar levels of variability were observed in all populations with a mean number of alleles per locus ranging from 10.3 to 11.6 and average observed heterozygosity across all loci ranging from 0.72 to 0.81.

At the population level, slight deviations from Hardy-Weinberg expectations occurred. They remained significant in three cases after the sequential Bonferroni procedure was applied over loci and populations (locus AFND2 in the population of Ngaoundere and Maga, locus FunD in Foumban). The Hardy-Weinberg disequilibrium across all loci remained significant only in one population, Foumban, and was due to one locus (FunD; Table 3). Thus, no major departure from Hardy-Weinberg equilibrium was evident in any population and in particular none matched the deficiency of heterokaryotypes in Tibati. Exact tests for linkage disequilibrium within each population (450 pairwise comparisons) resulted in four significant values after correction by the sequential Bonferroni procedure, each in different populations. Several loci are relatively close to one another in the genome (AF20, AF19, FunD, and FunG on arm 3R; see Figure 2); however, our results suggest absence of statistical linkage. Together with Hardy-Weinberg equilibrium, linkage disequilibrium analysis revealed that each geographical population, including Tibati, might be considered as panmictic.

Linkage disequilibrium between microsatellite loci and chromosomal inversion systems was assessed considering only specimens for which both microsatellite genotype and karyotype was known (78 individuals). Among 40 pairwise linkage disequilibrium tests, 3 were significant ($P < 0.05$) at the single test level. None of them remained significant after application of the Bonferroni correction, showing no association between chromosomal inversions and microsatellite alleles (Table 2).

Pairwise $F_{ST}$ estimates between populations and across all loci are shown in Table 4, together with corresponding geographical distance between sampling sites. Aver-
## Table 2
Pairwise linkage disequilibrium for 10 microsatellite loci and four chromosomal inversions for pooled *An. funestus* populations

<table>
<thead>
<tr>
<th>Microsatellite loci</th>
<th>Within inversions</th>
<th>Outside inversions</th>
<th>Not located</th>
<th>Chromosomal inversions</th>
</tr>
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<td>FunD</td>
<td>FunF</td>
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<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>FunG</td>
<td>-</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>FunD</td>
<td>-</td>
<td>+</td>
<td></td>
<td></td>
</tr>
<tr>
<td>FunF</td>
<td>-</td>
<td>-</td>
<td>+</td>
<td></td>
</tr>
<tr>
<td>Outside inversions</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>FunO</td>
<td>+</td>
<td></td>
<td>-</td>
<td>+</td>
</tr>
<tr>
<td>AFND20</td>
<td>-</td>
<td>-</td>
<td>+</td>
<td>-</td>
</tr>
<tr>
<td>AFND19</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>FunL</td>
<td>-</td>
<td>+&lt;sup&gt;a&lt;/sup&gt;</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Not located</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>AFND2</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>+</td>
</tr>
<tr>
<td>AFND3</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Chromosomal inversions</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>2R&lt;sub&gt;dha&lt;/sub&gt;</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>3R&lt;sub&gt;a&lt;/sub&gt;</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>3R&lt;sub&gt;b&lt;/sub&gt;</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>3L&lt;sub&gt;a&lt;/sub&gt;</td>
<td>+</td>
<td>-</td>
<td>-</td>
<td>+</td>
</tr>
</tbody>
</table>

*+, significant (P < 0.05) linkage disequilibrium before Bonferroni procedure; −, lack of significance (P < 0.05 level). 

<sup>a</sup>Significant (P < 0.05) linkage disequilibrium after Bonferroni procedure.
### Table 3

**Estimate of \( F_{IS} \) and significance level for goodness-of-fit tests to Hardy-Weinberg equilibrium for 10 microsatellite loci within *A. funestus* populations from 10 localities in Cameroon**

<table>
<thead>
<tr>
<th>Locus</th>
<th>Mfou (2n = 108)</th>
<th>Mbebe (2n = 112)</th>
<th>Ntui (2n = 104)</th>
<th>Nkoteng (2n = 112)</th>
<th>Foumban (2n = 96)</th>
<th>Bankim (2n = 96)</th>
<th>Tibati (2n = 176)</th>
<th>Ngaoundere (2n = 108)</th>
<th>Lagdo (2n = 146)</th>
<th>Maga (2n = 108)</th>
<th>Pooled* (2n = 1166)</th>
</tr>
</thead>
<tbody>
<tr>
<td>AFND2</td>
<td>-0.044</td>
<td>0.016</td>
<td>-0.022</td>
<td>0.061</td>
<td>0.017</td>
<td>0.028</td>
<td>0.043*</td>
<td>0.115**</td>
<td>-0.050*</td>
<td>0.111**</td>
<td>0.061**</td>
</tr>
<tr>
<td>AFND3</td>
<td>-0.025</td>
<td>0.004</td>
<td>0.241</td>
<td>-0.049</td>
<td>-0.011</td>
<td>-0.081</td>
<td>-0.148</td>
<td>-0.244</td>
<td>0.186</td>
<td>-0.074*</td>
<td>-0.024*</td>
</tr>
<tr>
<td>AFND5</td>
<td>0.059</td>
<td>0.031</td>
<td>0.068</td>
<td>-0.170</td>
<td>-0.114</td>
<td>-0.265</td>
<td>-0.075</td>
<td>0.133</td>
<td>-0.029</td>
<td>-0.033</td>
<td>-0.035</td>
</tr>
<tr>
<td>FunO</td>
<td>0.066</td>
<td>0.111</td>
<td>0.151</td>
<td>0.116</td>
<td>0.068</td>
<td>-0.043*</td>
<td>-0.015</td>
<td>0.003</td>
<td>-0.023*</td>
<td>0.008</td>
<td>0.049*</td>
</tr>
<tr>
<td>AFND20</td>
<td>0.168*</td>
<td>0.224*</td>
<td>-0.011</td>
<td>0.061</td>
<td>0.055</td>
<td>0.104</td>
<td>0.131*</td>
<td>0.076</td>
<td>0.016</td>
<td>0.128</td>
<td>0.097**</td>
</tr>
<tr>
<td>AFND19</td>
<td>0.099</td>
<td>0.013</td>
<td>0.030</td>
<td>-0.021</td>
<td>-0.068*</td>
<td>-0.023</td>
<td>0.039</td>
<td>0.062</td>
<td>0.146*</td>
<td>-0.028</td>
<td>0.029*</td>
</tr>
<tr>
<td>FunG</td>
<td>0.030</td>
<td>-0.009</td>
<td>0.130</td>
<td>0.041</td>
<td>-0.032</td>
<td>-0.038</td>
<td>-0.063</td>
<td>-0.122</td>
<td>-0.085</td>
<td>0.020</td>
<td>0.000</td>
</tr>
<tr>
<td>FunD</td>
<td>0.089</td>
<td>0.000</td>
<td>0.065</td>
<td>-0.005</td>
<td>0.078**</td>
<td>-0.005</td>
<td>-0.009</td>
<td>0.058*</td>
<td>-0.007</td>
<td>0.042</td>
<td>0.051**</td>
</tr>
<tr>
<td>FunL</td>
<td>0.036</td>
<td>0.005</td>
<td>-0.045</td>
<td>-0.010</td>
<td>0.080</td>
<td>0.070</td>
<td>0.108</td>
<td>0.008</td>
<td>-0.049</td>
<td>0.050</td>
<td>0.029</td>
</tr>
<tr>
<td>FunF</td>
<td>-0.035</td>
<td>-0.005</td>
<td>-0.112</td>
<td>-0.012</td>
<td>0.200</td>
<td>0.020</td>
<td>-0.022</td>
<td>-0.072</td>
<td>-0.022</td>
<td>-0.068</td>
<td>-0.003</td>
</tr>
<tr>
<td>Mean across all loci*</td>
<td>0.044</td>
<td>0.039</td>
<td>0.065</td>
<td>-0.012</td>
<td>0.012</td>
<td>0.027*</td>
<td>-0.016*</td>
<td>-0.010</td>
<td>-0.011*</td>
<td>0.009*</td>
<td>0.016</td>
</tr>
</tbody>
</table>

\( F_{IS} \) was calculated according to Weir and Cockerham (1984). 2n, number of chromosomes scored. *P < 0.05; **P < 0.05 after Bonferroni correction (see text).

* All specimens were considered as belonging to one single panmictic population.

* Multilocus \( F_{IS} \) estimates were computed according to Weir and Cockerham (1984) using GENEPOP 3.2. (Raymond and Rousset 1995); statistical significance of \( F_{IS} \) across loci was assessed through Fisher’s exact test.
Measure of genetic differentiation ($F_{ST}$) and geographical distance among An. funestus populations from Cameroon

<table>
<thead>
<tr>
<th></th>
<th>Mfou</th>
<th>Mbebe</th>
<th>Ntui</th>
<th>Nkoteng</th>
<th>Foumban</th>
<th>Bankim</th>
<th>Tibati</th>
<th>Ngaoundere</th>
<th>Lagdo</th>
<th>Maga</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mfou</td>
<td>—</td>
<td>80</td>
<td>85</td>
<td>105</td>
<td>250</td>
<td>270</td>
<td>330</td>
<td>460</td>
<td>640</td>
<td>880</td>
</tr>
<tr>
<td>Mbebe</td>
<td>0.0062*</td>
<td>—</td>
<td>50</td>
<td>100</td>
<td>170</td>
<td>190</td>
<td>290</td>
<td>420</td>
<td>580</td>
<td>830</td>
</tr>
<tr>
<td>Ntui</td>
<td>0.0083*</td>
<td>0.0071</td>
<td>—</td>
<td>50</td>
<td>195</td>
<td>175</td>
<td>250</td>
<td>380</td>
<td>560</td>
<td>800</td>
</tr>
<tr>
<td>Nkoteng</td>
<td>0.0002*</td>
<td>—</td>
<td>0.0022</td>
<td>—</td>
<td>0.0037</td>
<td>—</td>
<td>190</td>
<td>180</td>
<td>230</td>
<td>380</td>
</tr>
<tr>
<td>Foumban</td>
<td>0.0164*</td>
<td>0.0289*</td>
<td>0.0184*</td>
<td>0.0203*</td>
<td>—</td>
<td>80</td>
<td>220</td>
<td>360</td>
<td>490</td>
<td>740</td>
</tr>
<tr>
<td>Bankim</td>
<td>0.0107*</td>
<td>0.0137*</td>
<td>0.0026</td>
<td>0.0099*</td>
<td>0.0095*</td>
<td>—</td>
<td>130</td>
<td>260</td>
<td>410</td>
<td>660</td>
</tr>
<tr>
<td>Tibati</td>
<td>0.0203*</td>
<td>0.0295*</td>
<td>0.0123*</td>
<td>0.0199*</td>
<td>0.0079*</td>
<td>0.0044*</td>
<td>—</td>
<td>150</td>
<td>310</td>
<td>540</td>
</tr>
<tr>
<td>Ngaoundere</td>
<td>0.0350*</td>
<td>0.0386*</td>
<td>0.0165*</td>
<td>0.0286*</td>
<td>0.0231*</td>
<td>0.0113*</td>
<td>0.0046*</td>
<td>—</td>
<td>190</td>
<td>410</td>
</tr>
<tr>
<td>Lagdo</td>
<td>0.0281*</td>
<td>0.0368*</td>
<td>0.0146*</td>
<td>0.0250*</td>
<td>0.0315*</td>
<td>0.0136*</td>
<td>0.0108*</td>
<td>0.0112</td>
<td>—</td>
<td>240</td>
</tr>
<tr>
<td>Maga</td>
<td>0.0298*</td>
<td>0.0373*</td>
<td>0.0145*</td>
<td>0.0283*</td>
<td>0.0224*</td>
<td>0.0102*</td>
<td>0.0028*</td>
<td>0.0015</td>
<td>0.0067*</td>
<td>—</td>
</tr>
</tbody>
</table>

$F_{ST}$ is below the diagonal. Distance in kilometers is above the diagonal. *$P < 0.01$.

Mean $F_{ST}$ estimates were computed separately for loci within (AFND5, 0.0066; FunG, 0.0212; FunD, 0.0267; FunF: 0.0090) and outside (FunO, 0.0169; AFND20, 0.0018; AFND19, 0.0150; FunL, 0.0014) chromosomal inversions. Comparison by Mann-Whitney test was non-significant ($P > 0.05$) but power was clearly low, which suggests homogeneity among these groups of loci and negligible (if any) influence of chromosomal inversions on our estimates.

Isolation by distance was tested and statistically significant correlation between genetic (pairwise $F_{ST}$) and geographic distance ($P < 10^{-4}$) was detected when considering the whole data set (Figure 3). These results suggest that the level of genetic differentiation between populations of An. funestus in Cameroon is due to a decrease of gene flow by geographical distance. To investigate whether one or few loci were responsible for the significant results, we repeated the analysis by keeping, in a stepwise procedure, all data except for one locus. Then we considered only loci located within chromosomal inversions or only loci located outside inversions. The results remained significant regardless of which locus was removed or which group of loci was considered ($P < 0.05$). These data suggest that isolation by distance is due to the whole set of loci.

**DISCUSSION**

Adaptive role of chromosomal inversions: In An. funestus populations from Cameroon, chromosomal inversions were distributed along a cline of frequencies. Similar clines in frequencies of chromosomal inversions associated with an environmental gradient were previously observed in other Diptera. In the Drosophila genus, paracentric inversions are almost ubiquitous and their distribution was found to correlate with contrasting environmental conditions (Anderson 1989; Krimbas and Powell 2000). Such a distribution was explained by the fact that chromosomal inversions would be maintained through directional selection pressure. In African malaria vectors, the biological significance of inversion polymorphism has been studied in depth in two species of the An. gambiae complex, namely An. gambiae s.s. and An. arabiensis. Chromosomal inversions were also found to be distributed according to ecotypic conditions (Coluzzi et al. 1979, 1985; Coluzzi 1982; Toure et al. 1998; Petrarca et al. 2000) and are supposed to be strongly exposed to...
environmental selection (Coluzzi et al. 1979). In An. funestus populations from Cameroon, the clines of frequencies of chromosomal inversions along the transect, together with strong linkage disequilibrium between inversions located on distinct chromosomes, are therefore in agreement with a similar adaptive role of inversions: 2Rd, 2Rh, 2Ra, 3Ra, 3Rh, and 3La would favor adaptation to moist equatorial climates, while standard arrangements would confer a better adaptive value under drier conditions.

The taxonomic status of chromosomal forms in An. funestus—reconciling microsatellite and chromosomal data: According to the speciation hypothesis proposed by Costantini et al. (1999), both Folonzo and Kiribina chromosomal forms are considered isolated reproductive units, each one having its own set of chromosomal inversions. Thus, although certain chromosomal inversions may be shared between them, others might have diagnostic values and could be considered as specific to one or the other of these forms. This is the case, for example, between An. gambiae and An. arabiensis, both established species being characterized by fixed inversions on chromosome X, and with inversion 2La fixed in An. arabiensis and polymorphic in An. gambiae (Toure et al. 1998; Coluzzi et al. 2002). In this case, evidence for genetic divergence between species/populations should be found throughout the genome, the amplitude of which will depend on time since lineage splitting. High amounts of genetic differentiation were indeed revealed between An. gambiae and An. arabiensis using, among others, microsatellite markers (Kamau et al. 1998; Lanzaro et al. 1998; Wang et al. 2001; Besansky et al. 2003). Our data are in poor agreement with this hypothesis in An. funestus.

In the forest-savanna transition area where chromosomal inversions were polymorphic, we found two contrasting patterns. In Tibati, chromosomal structuring suggested local subdivision of An. funestus in chromosomal forms. This is reminiscent of the situation in West Africa, which led to characterization of the Folonzo and Kiribina forms (Lochouarn et al. 1998; Costantini et al. 1999; Dia et al. 2000a). In Bankim, however, panmixia was not rejected. The same situation was formerly observed in Angola (Boccolini et al. 2002), Kenya (Sharakhov et al. 2001; Kamau et al. 2003), and Madagascar (G. Le Goff, personal communication), thus underlining that the subdivision in chromosomal forms could not be generalized to the whole species range.

Microsatellite markers showed no departure from panmixia in any geographical population from Cameroon. Microsatellite loci located within polymorphic chromosomal inversions, as well as loci located outside, detected isolation by distance between geographical isolates, thus providing support for the usefulness of microsatellite markers to detect population substructure in An. funestus and negligible, if any, effect of allele size homoplasy. Recombination between inverted chromosomal regions might occur in heterozygous individuals through multiple crossing over or gene conversion. Although such a phenomenon is expected to occur at low frequency, it has been shown that recombination rate and gene flow are highly site dependent inside inverted regions: they are maximal in the central region of the inversion and minimal at the breakpoints (Navarro et al. 1997; Andolfatto et al. 2001). They also depend on the length of the inversions and on the species studied (Caceres et al. 1999). It is therefore likely that the regions examined by our microsatellite markers are not the most affected by the reduction of recombination, even if some of them are close to the breakpoints (i.e., FunG and AFND5). High-resolution mapping using additional microsatellite and/or single nucleotide polymorphism markers should help pinpoint chromosomal regions, as well as the genes they contain, that are most exposed to environmental selection and potentially involved in ecotypic adaptation.

Altogether, our results are inconsistent with complete reproductive isolation between chromosomal forms. The conundrum in our data is that inversion heterozygotes apparently are inviable or do not occur frequently, and yet there is no allelic differentiation of markers found within inversions. The apparent homogeneity of the gene pool might result from historical rather than current gene flow between chromosomal forms. This is an issue that warrants further investigation.

Inversion heterozygote deficiency: Positive assortative mating, i.e., the general trend for like inversions to mate together more often than expected if mating is random, could explain the excess of homozygous karyotypes observed within An. funestus populations. Assortative mating could be due to mate choice favoring mating between individuals belonging to the same chromosomal form, or chromosomal forms could be separated temporally or spatially at the microgeographic scale (i.e., parapatry) at the time mating occurs, although sympathy is observed at the adult stage when sampling blood-seeking or resting females. However, this hypothesis cannot explain the pattern observed at the continental scale, with an inversion subdivision of An. funestus populations in West Africa while chromosomal inversions reveal no heterokaryotype deficit in South and East Africa.

Another hypothesis that fits better with the pattern of observed data involves selection acting against hybrid heterokaryotypes. An. funestus chromosomal forms may freely interbreed in nature, but strong underdominance due to environmental selection against hybrid heterokaryotypes would limit their development. As we found both chromosomal forms to be represented at the adult stage when sampling blood-seeking and resting females, we assume that differential environmental adaptation between chromosomal forms would play a role at earlier stages. Selection pressure on chromosomal inversions could therefore occur at the larval stage, with different types of breeding sites being preferred by each chromo-
The control of malaria in Africa: Our study deals with one of the most efficient human malaria vectors in the world, which despite its epidemiological importance in malaria transmission (Gillies and de Meillon 1968) has been widely overlooked by the scientific community. This may be partially attributed to the fact that An. funestus is much harder to breed in the laboratory than An. gambiae, thereby hindering classical genetic mapping and transformation experiments. However, if the ultimate goal of studies of malaria vectors is to eventually achieve control of the disease in the field, this deficiency is not acceptable. Any population genetic control measure to be implemented, such as autocidal control (see Gould and Schliekelman 2004) or introduction of transgenes for Plasmodium resistance, needs to account for the presence in the field of numerous vector species and populations among which gene flow appears to be restricted or not. In the case of An. gambiae, gene flow has been observed to be restricted by inversion (Triquet et al. 2004), but in the case of An. funestus in Cameroon, inversions do not restrict gene flow. Thus, inversions could interfere with the spread of transgenes in An. gambiae, but appear to be of less concern for An. funestus whenever transformation is available for this species. The success of any strategy for malaria control in Africa through genetic modification of its mosquito vectors will rely on our ability to target all the species and populations that significantly contribute to the transmission and will require a thorough understanding of the relevant population genetics.

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LITERATURE CITED


Fontenille, D., L. Lochoeur, N. Dramo, C. Sokhina, J. J. Lemasson et al., 1997 High annual and seasonal variations in malaria